



ATLAS Urine Reagent Strips

6 Parameters

Glucose, Protein, Nitrite, pH, Blood, and Leukocytes in urine.

For *In-Vitro* and professional use only

Atlas urine reagent strips are intended for professional use. The "universal precautions" recommended by the Centers for Disease control should be adhered to whenever blood or body fluids are handled.¹

These precautions include wearing gloves. ATLAS urine reagent strips may contain either diazonium salt or nitroferrocyanide. Avoid contact with skin and mucous membranes; flush affected areas with copious amounts of water. Get immediate medical attention for eyes or if ingested. Exercise the normal precautions required for handling all laboratory reagents.

SUMMARY AND INTENDED USE:

ATLAS urine reagent strip is a dip-and-read test strip and intended for use as an in vitro diagnostic and using urine specimens. The strip contains solid phase reagent areas affixed to a plastic support and is provided in a dry reagent format. This strip provides qualitative and semi quantitative tests for, **glucose, protein, nitrite, pH, occult blood, and leukocytes** by the visual comparison with color charts of each concentration range. No additional reagents or laboratory equipment is required. The reagent strips are packaged in a plastic vial containing desiccant. The test strips must be tightly capped in the plastic vial to assure reagent reactivity. The directions must be followed exactly, and it is necessary to use fresh, well-mixed and uncentrifuged urine for optimal results.

CHEMICAL PRINCIPLES OF THE PROCEDURE:

Glucose: This test is based on a sequential enzyme reaction. First, glucose oxidase catalyzes the formation of gluconic acid and hydrogen peroxide from the oxidation of glucose. A second enzyme, peroxidase, catalyzes the reaction of hydrogen peroxide with potassium iodide chromogen to oxidize the chromogen to colors ranging from blue through greenish-brown to dark brown.

Protein: This test is based on the color change of the indicator, tetrabromophenol blue, in the presence of protein. A positive reaction is indicated by a color change from yellow through green and then to greenish-blue.

Nitrite: This test is based on the reaction of p-arsanilic acid and nitrite (which is derived from dietary nitrate in the presence of bacteria) in urine to form a diazonium compound. The diazonium compound in turn couples with N-(1-naphthyl)ethylenediamine in an acidic medium. The resulting color is pink. Any degree of pink color is considered positive.

pH: This test is based on double indicators (methyl red and bromothymol blue) which give a broad range of colors covering the entire urinary pH range. Colors range from orange through greenish-yellow and green to blue.

Blood: This test is based on the pseudoperoxidase activity of hemoglobin which catalyzes the reaction of 3,3',5,5'-tetramethylbenzidine and buffered organic peroxide, 2,5-dimethylhexane-2,5-dihydroperoxide. The resulting color ranges from, greenish-yellow through bluish-green to dark blue.

Leukocytes: This test reveals the presence of granulocyte esterases. The esterases cleave a derivatized thiazole amino acid ester to liberate derivatized hydroxy thiazole. This thiazole then reacts with a diazonium salt to produce a purple product.

REAGENTS: (Based on dry weight at time of impregnation of 100strips)

Glucose	Glucose oxidase	451unit
	Peroxidase	186unit
	Potassiumiodide	10.0mg
Bilirubin	2,4-Dichlorobenzenediazonium Oxalic acid	3.0mg
	Protein	30.0mg
Nitrite	Tetrabromophenol blue	0.3mg
	Citric acid	110.0mg
	Trisodium citrate	46.0mg
pH	p-Arsanilic acid	5.0mg
	N-(1-naphthyl)ethylenediamine	6.0mg
	Methyl red	0.04mg
Blood	Bromothymol blue	0.5mg
	2,5-Dimethylhexane-2,5dihydroperoxide	40.0mg
	3,3'5,5'- Tetramethylbenzidine	3.7mg
Leukocytes	2-Phenylthiazole amino acid ester	1.0mg
	Diazonium salt	0.7mg

STORAGE:

Store at room temperature between 15°C-30°C (59°F-86°F). Do not store the strips in the refrigerator or freezer. Cap the bottle tightly. Since the test strips are sensitive to specific environmental factors, such as moisture, heat and light, do not expose strips to these factors.

PROCEDURE FOR HANDLING THE STRIPS:

All unused strips must be stored in the original bottle. Do not remove desiccant from bottle. Transfer of the strips to another container may cause reagent strips to deteriorate and become uncreative. After taking out test strips, replace the cap, promptly and tightly. Do not touch test area of the strip. Do not use strips after expiration date. The work area should be clean and free of detergents and other contaminants.

SPECIMEN COLLECTION AND PREPARATION:

Use a clean, dry, unused vessel to collect the urine. Test the urine as soon as possible after collection. If testing cannot be done within an hour after voiding, refrigerate the specimen immediately and let it return to room temperature before testing.

PROCEDURE:

This procedure MUST BE FOLLOWED EXACTLY to achieve reliable test results.

1. Confirm that the product is within the expiration date shown on the label.
2. Remove the strip from the bottle and replace the cap immediately.
3. Inspect the strip. Discoloration or darkening of reagent areas may indicate deterioration. Do not use strip.
4. Dip the test strip completely for no more than 1 second in fresh, well-mixed, and uncentrifuged urine specimen. Excessive urine on the test strip may give rise to a wrong result. Remove the excessive urine by touching the plastic film on the rim of vessel. At this time, do not allow the reagent areas to touch the rim of vessel. Excessive urine may be removed by gently blotting the lengthwise edge on absorbent paper.
5. Compare the test results carefully with the color chart on the bottle label in good light. Proper reading time (30-60 seconds) is critical for optimal results. While comparing, keep the strip in a horizontal position to avoid possible interaction of chemicals by excessive urine. Changes in color that appears only along the edges of the test areas or after the correct timing period has passed are of no diagnostic significance.

QUALITY CONTROL:

The strips must be properly stored and handled before and during the testing. Reaction of reagent strips should be confirmed by testing known positive and negative specimens or multiple analyte controls containing normal and abnormal amounts of each of the analytes being tested.

RESULTS:

The results are obtained by direct comparison of test strip with the color blocks printed on the bottle label. No calculations or laboratory instruments are necessary.

LIMITATIONS OF PROCEDURE:

Substances that cause abnormal urine color, such as drugs containing azo dyes, nitrofurantoin and riboflavin may affect the readability of reagent areas on urinalysis reagent strips. The color development on the reagent pad may be masked, or a color reaction may be produced on the pad that could be interpreted visually and instrumentally as a false positive. It is therefore recommended that in case of doubt, the test should be repeated after withdrawal of the medication.

Glucose: Reactivity of the test decreases as the specific gravity and/or pH of urine increases, and may also vary with temperature. Ascorbic acid (more than 50mg/dl) and ketone bodies (more than 40mg/dl) may cause a false negative for a specimen containing a small amount of glucose (100mg/dl), however, the combinations of such ketone levels and low glucose levels are metabolically improbable in screening.

Bilirubin: Normally no bilirubin is detectable in urine by even the most sensitive methods. Since the bilirubin in specimens is sensitive to light, exposure of the urine specimens to light for a long period time may result in a false negative.

Ascorbic acid concentration of 25-50mg/dl may also cause false negatives. Even trace amounts of bilirubin are sufficiently abnormal to require further investigation. False positive results may be obtained in the presence of diagnostic or therapeutic dyes in the test urine.

Protein: The minimum sensitivity of this test is 10-20mg/dl of protein in urine. Highly buffered alkaline urine (pH 9) may give false negative results. The interpretation of results is also difficult in turbid urine specimens.

Nitrite: Any degree of uniform pink color development should be considered positive, however, pink spots or pink edges should not be interpreted as a positive result. Color development is not proportional to the number of bacteria present. The urine test detects only nitrate reducing bacteria. Occasionally bacteria will be present that do not reduce nitrate to nitrite. Therefore, a negative result cannot rule out the presence of urinary tract infection or bacteriuria. A mid stream of first morning urine specimen is recommended for this test.²Sensitivity of the nitrite test is decreased with high specific gravity or ascorbic acid concentrations of 25mg/dl or greater.

pH : This pH test indicates the pH values only within the range of 5 to 9. Certain drugs, such as those used for hypertension and heart trouble (Acetazolamides) may cause alkaline urine.²Excessive urine on the test strip may wash the acid buffer from the neighboring protein reagent onto the pH area and change the pH reading to an acid pH although the urine being tested is originally neutral or alkaline. This is called the "run-over" phenomenon.

Blood: A false positive can sometimes occur when bacteria are present in the urine. Ascorbic acid or protein may reduce the reactivity of the blood test. Strong oxidizing substances, such as hypochlorites, may produce a false positive result. Urine from menstruating females often, but not always, yields positive results.

Leukocytes: The test result may not always be consistent with the leukocyte cell number by the microscopic examination. Positive results may be found in high humidity and high temperature condition, and failure of the bottle security. Positive results may occasionally be found with the random specimens from females due to contamination of the specimens by the vaginal discharge. High concentration of glucose, high specific gravity, high level of albumin, high concentration of formaldehyde, or presence of blood may cause decreased test results. High concentration of oxalic acid or trace of oxidizing agents may cause false negative results. Low specific gravity (1.010 and under) or pH 6.5 ≥ may cause a false positive.

EXPECTED VALUES:

Glucose: Normally no glucose is detectable in urine, although the normal kidney excretes a minute quantity of glucose. Approximately 100mg glucose/dl urine is detectable with this test strip. Concentrations of 100mg/dl may be considered as abnormal if found consistently.

Ketones: Ketone bodies should not be detected in normal urine specimens with this reagent. The concentration given : ±(5mg/dl), +(15mg/dl), ++(40mg/dl), +++(80mg/dl) correlate well with the acetoacetic acid concentration in urine. The sensitivity of this test is 5mg acetoacetic acid per 100ml of urine. Detectable levels of ketone may occur with frequent vomiting, diarrhea, digestive disturbances, pregnancy, or severe physical exercise.

Protein: Normal urine specimens ordinarily contain some protein (0-4mg/dl); therefore, only persistent elevated levels of urine protein indicate kidney or urinary tract disease. The persistent result of trace level or over indicates significant proteinuria, and thus further clinical testing is needed to evaluate the significance of results. The concentrations given; "+"(30mg/dl); "++"(100mg/dl); "+++"(300mg/dl); "++++"(2000mg/dl or more); correlate well with the albumin concentrations in urine. Pathologic proteinuria generally gives values above 30mg/dl and is persistent.

Nitrite: Nitrite indicates the presence of bacteria in urine. Any degree of pink color after 30 seconds indicates clinically significant bacteriuria. Bacteriuria is generally due to infection of the kidneys, ureters, bladder or urethra.

pH: Normal urine is slightly acid with a pH of 6 and urine pH values generally range from 5 to 8 pH units. The pH of urine is an important indicator of certain metabolic, kidney, gastrointestinal and respiratory factors.

Blood: Hemolysis is a natural process of recycling old or damaged red cells. But when hemoglobin appears in urine, it indicates some kind of kidney disease or some kind of urinary

tract disorder. The practical detection limit of this test is approximately 5 to 10 erythrocytes per microliter of urine. Blood is often, but not always, found in the urine of menstruating females. This test is highly sensitive to hemoglobin (it is slightly less so to intact erythrocytes) and thus complements the microscopic examination.

Leukocytes: Normally no leukocytes are detectable in urine. Individually observed trace results may be questionable clinical significant.

PERFORMANCE CHARACTERISTICS:

Specific performance characteristics of ATLAS products are based both on clinical and laboratory studies. A study done at two clinical sites involving 94 patient samples compared ATLAS urine reagent strip to a competitor's strip. 100% agreement within one color block was obtained for all analytes except protein. Protein gave a greater than 95% agreement. The lower agreement may be reflective of the technician's interpretation of the negative versus trace color block with both ATLAS urine reagent strips and the competitive strip. Parameters of importance to the user are sensitivity, limits of test, specificity, accuracy, precision and stability. Sensitivity and limits of tests are the generally detectable levels of each test described previously. The sensitivity depends upon several factors; the variability of color perception; the presence or absence of inhibitory factors typically found in urine, the specific gravity, ascorbic acid and pH; and lighting conditions when the product is read visually. The tests have been developed to be specific for the constituent to be measured with the exception of interferences listed previously. Exact agreement between visual results and instrumental results might not be found because of the inherent differences between the perception of the human eye and the optical system of the instruments. It is for this reason that each user is encouraged to develop his own standards for performance. The stability test has been developed by statistical procedure for various environmental conditions.

Glucose: The test has a sensitivity of 100mg glucose in 100ml urine and is specific for glucose. No substance excreted in urine other than glucose is known to give a positive result. False negative results may be obtained with the presence of levodopa, ascorbic acid, glutathione, and dipyrone. If the test color appears some what mottled at the higher glucose concentrations, match the darkest color-to-color blocks.

Bilirubin: The test has a sensitivity of 0.5mg/dl bilirubin. Bilirubin in urine indicates liver disease before any clinical signs are usually evident.

Protein: The test is more sensitive to albumins than to gamma-globulins. Bence-Jones proteins, and mucoproteins ; such proteins do not interfere with the reaction of albumin.

Nitrite: This test has a sensitivity of 0.05mg/dl nitrite ion in urine of normal specific gravity and moderate levels of ascorbic acid. The test is specific for nitrite and independent of urinary pH of any other substance normally excreted in

urine. Comparison of the reacted area against a white background may aid in the detection of low levels of nitrite.

pH: This test produces distinct color changes from orange to blue over the pH value 5-9. Values will be read to within 1 unit period. Slight variations may be caused by the pigments in the urine.

Blood: This test is slightly more sensitive to free hemoglobin and myoglobin than to intact erythrocytes. The test is generally capable of detecting 0.015mg/dl free hemoglobin or 5 to 10 intact red blood cells per microliter urine. The sensitivity may be reduced in urine specimens with high specific gravity and ascorbic acid content. The appearance of green spots on the reagent test area indicates the presence of intact erythrocytes in the urine.

Leukocytes: The test is generally capable of detecting 20-25WBC/ μ l as a trace.

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